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Protective effect of chaperones on polyglutamine diseases

Yasushi Kobayashi and Gen Sobue*

Department of Neurology, Nagoya University Graduate School of Medicine, Nagoya, Japan

ABSTRACT: Polyglutamine diseases are inherited neurodegenerative diseases caused by the expansion of polyglutamine tract in the disease causing gene products. Studies of polyglutamine disease patients and transgenic mice have revealed that nuclear inclusions formed by the disease protein are a common pathological feature of these diseases. The finding that nuclear inclusions are ubiquitinated raises the possibility that alterations in the major intracellular system for degrading proteins, the ubiquitin-proteasome pathway, may be involved in the pathogenesis of polyglutamine diseases. Perturbations in proteasome function are associated with altered expression levels of stress-response or heat shock proteins. Heat shock proteins function as molecular chaperones, which recognize and renaturate misfolded protein (aggregate). In this article, we review the role of chaperones in the development of polyglutamine diseases. Overexpression of chaperones reduces aggregate formation and suppresses apoptosis in several polyglutamine disease models including spinal and bulbar muscular atrophy. These facts indicate that chaperones may be one of the key factors in the development of polyglutamine disease, and suggest that increasing expression level or enhancing the function of chaperones will provide an avenue for the treatment of polyglutamine disease. © 2001 Elsevier Science Inc.

KEY WORDS: Chaperone, Polyglutamine, Hsp70, Hsp40, SBMA.

INTRODUCTION

Polyglutamine diseases are inherited neurodegenerative diseases caused by the expansion of CAG repeats (polyQ) in the disease causing genes [55]. For example, in spinal and bulbar muscular atrophy (SBMA) patients, a normally polymorphic CAG repeats (10–36 CAGs) expands to 38–66 CAGs in the first exon of *androgen receptor* gene. The number of CAGs is inversely correlated with the age of onset of the disease [15,32]. To date, several polyQ diseases have been identified, including SBMA, Huntington's disease (HD) [50], dentatorubralpallidoluysian atrophy [31, 41], Machado-Joseph disease (MJD) [27], and others. The number of identified polyQ diseases is still increasing. The polyQ diseases have different disease-causing genes, suggesting that these disorders share a common pathogenesis involving the gain of a toxic function associated with the expanded polyglutamine tract.

Processing of the polyglutamine-containing disease protein by proteases (e.g., caspase family) may liberate truncated fragments with the polyglutamine tract [17,30]. Truncated proteins with the expanded polyglutamine tracts cause neurodegeneration in transgenic mice as well as *Drosophila*, and cell death in transfected cells [13,22,38,52]. In addition to cellular toxicity, truncated proteins with the expanded polyglutamine tracts have been shown to form aggregates, likely through hydrogen bonding or transglutaminase activity [25,44,49]. Studies of polyQ disease patients and transgenic mice have revealed that nuclear inclusions (NI) formed by the disease protein are a common pathological feature of these diseases [14,21,33,34,43,47]. In SBMA, NIs containing androgen receptor (AR) protein have been mainly observed in the regions of SBMA central nervous system susceptible to degeneration, including the brain stem motor nuclei and spinal motor neurons [33,34]. The finding that NIs are ubiquitinated raises the possibility that alterations in the major intracellular system for degrading proteins, the ubiquitin-proteasome pathway, may be involved in the pathogenesis of polyQ diseases. The proteasome is a large multicatalytic protease complex that is critical for many cellular processes including cell cycle control, differentiation, antigen presentation, and cell survival [11]. Perturbations in proteasome function are associated with altered expression levels of stress-response or heat shock proteins (Hsps) [2]. These proteins function as molecular chaperones, which recognize and renaturate misfolded proteins under normal and stressed conditions. In addition, chaperones may maintain proteins in an appropriate conformation [20]. The possible role of chaperones in the development of polyQ diseases is raising considerable interest.

HSP70 AND HSP40 AS MOLECULAR CHAPERONES

It has been postulated that most Hsps have a molecular chaperone activity involved in various aspects of protein metabolism [20]. A molecular chaperone is a substance that binds to a substrate protein irrespective of stability, and facilitates its fate in the right way *in vivo*, i.e., folding, oligomeric assembly (switching active/ inactive conformations), or transport to a subcellular compartment. The mechanism of this function is to control binding and releasing the substrate proteins [16]. Hsp70 and the Hsp40 chaperone family members act together as molecular chaperones [4]. Hsp40 family regulates the chaperone activity of Hsp70 family through upregulation of their ATPase activity [19]. This chaperone complex system works ubiquitously from bacteria to mammals [4]. Hsp70 in cooperation with Hsp40 has been demonstrated to renaturate heat-induced aggregation *in vitro* and *in vivo* [39,40]. This function to renaturate aggregation is dependent on Hsp40-enhanced adenosine triphosphate hydrolysis [40]. Most Hsps, including Hsp105 [53], Hsp90 [6], Hsp60 [2], Hsp70/40 [39], and Hsp27 [36], work as molecular chaperones, and participate in various

^{*} Address for correspondence: Dr. Gen Sobue, Department of Neurology, Nagoya University Graduate School of Medicine, 65 Tsurumai-cho, Showa-ku, Nagoya 466-8550, Japan. Fax: +81-52-744-2384; E-mail: sobueg@med.nagoya-u.ac.jp

aspects of protein fate and, consequently, in the protection of living cells from deleterious environmental stresses.

CHAPERONE AND POLYQ DISEASES

Recently, overexpression of Hsps has been reported to decrease aggregate formation by expanded polyglutamine tract [12]. We hypothesized that the ability of Hsp70 and Hsp40 chaperones to facilitate refolding or proteolysis of mutant protein may be a key factor for neuronal cells to defend themselves against the toxic properties of expanded polyglutamine tract. Immunohistochemical studies revealed that polyQ-formed nuclear aggregates colocalized with several Hsps and proteasome $[7,12,48,54]$, however, naïve cellular Hsps and proteasomes could suppress neither aggregate formation nor cellular toxicity induced by polyQ. To determine whether a sufficient amount of chaperones could be effective to protect cells against toxicity of expanded polyglutamine tract, we overexpressed chaperones in the cell model of SBMA [29]. Our results reveal that a combination of Hsp70 and Hsp40, or Hsp70 alone, has a favorable effect on cellular protection as well as suppression of aggregate formation, and that combination of Hsp70 and Hsp40 has the strongest effect among them. Our results were in accordance with the reports that the chaperone function of Hsp70 is critically dependent on the cooperation with Hsp40 [4]. Hsps have been confirmed to suppress aggregate formation and cellular toxicity in other polyQ disease models [5,9,12,24,29]. In other conformational diseases, the study of mutant Cu/Zn-superoxide dismutase associated with familial form of amyotrophic lateral sclerosis also displayed that increasing the level of Hsp70 reduced formation of mutant SOD-containing aggregates in cultured primary motor neurons expressing mutant SOD-1 and prolonged their survival [1]. It is therefore reasonable to consider that disease gene product-formed aggregates are directly associated with the induction of neurodegeneration in polyQ disease and other conformation diseases. We thus reasoned that overexpression of both Hsp70 and Hsp40 chaperones reduces cytotoxicity induced by aggregate formation with disease gene product in polyQ disease and other conformation diseases.

The molecular mechanism for the reduction of cytotoxicity through inhibition of expanded polyglutamine tract formed-aggregate by overexpression of chaperones needs to be examined. Although it has been proposed that expanded polyglutamine tract formed-aggregates participate in inappropriate protein-protein interactions that lead to cell death, the nature of such interactions and the mechanism by which cell death is induced remain unclear. Molecular chaperones could be involved in the actual formation of expanded polyglutamine tract formed aggregates by stabilizing the unfolded protein in an intermediate conformation which has the propensity to interact with self or other proteins. To date, several proteins interacting with polyglutamine tract-containing disease gene product have been cloned, including huntingtin-associated protein [35], huntingtin-interacting protein [26], glyceraldehyde 3-phosphate dehydrogenase [3], leucine-rich acidic nuclear protein [37], PQBP-1 [51], and TAF130 [46]. These interacting proteins are candidate players in the pathogenesis of polyQ disease. Chaperones might reduce cytotoxicity of expanded polyglutamine tract through inhibition of the interaction of expanded polyglutamineformed aggregate with these proteins.

Another possibility is that overexpression of chaperones enhances the function of the ubiquitin-proteasome pathway for mutant protein degradation, because the function of the ubiquitinproteasome pathway is related with the expression level of chaperones [2]. Nuclear aggregates are ubiquitinated and are colocalized with chaperones and proteasome, implicating the ubiquitin-proteasome degradation pathway in the pathogenesis of polyQ disease [12,33,34]. The report that the inhibition of proteasome function accelerates aggregate formation by polyglutamine tract also implies that the ubiquitin-proteasome degradation pathway plays a direct role in modulating aggregation in polyQ disease [8]. In Alzheimer's disease, one of the conformational disease, amyloid protein, which is a major component of senile plaques, could interfere with ubiquitin-dependent protein degradation pathway by inhibiting the 26S proteasome. Consequently, this mechanism could lead to neuronal damage observed in Alzheimer's disease [18]. Similarly, expanded polyglutamine tract would interfere with ubiquitin-dependent protein degradation pathway and lead to neuronal damage in polyQ disease. Thus, overexpression of chaperones would enhance the function of proteasome, leading to the protection of cells expressing truncated and expanded AR against a cellular toxicity of expanded polyglutamine tract.

It has been recently reported that overexpression of Hsdj/Hdj2 in Hela cells decreases the frequency of mutant ataxin-1 and mutant AR aggregation [12,48]. However, we found that overexpression of Hsdj/Hdj2 has little effect on the reduction of aggregate formation and the protection from cytotoxicity in our report [29]. Such differences in the results may derive from the difference in cell lineage, the different origin of Hsdj/Hdj2 or expression level of Hsdj/Hdj2 in transfected cells. Previous studies used a nonneural cell line (Hela cell) [12,48], whereas we used a neural cell line (Neuro2a) [29]. The difference in cell lineages could influence the relations of chaperones, aggregation and cell death. Although Hsdj we employed in this study [29] is 99% identical to Hsdj2 employed in other reports [12,48] at the level of amino acid sequence [10,42], the relationship between Hsdj and Hdj2 remains to be studied.

In contrast to our results, evidence against a critical role of intranuclear aggregates in neuronal cell death has been provided [28,45]. Such discrepancies could arise from the difference of cell type/animal (primary neuron/animal vs. cell line) or interventions (neurotrophic factors/suppression of ubiquitin-conjugating enzyme/inhibition of ataxin-1 self-association vs. overexpression of chaperones). However, we cannot rule out the possibility that our finding of a parallel correlation between the reduction of aggregate formation and suppression of apoptosis in our system [29], independently might occur. Thus, Hsps could reduce aggregate formation as a molecular chaperone, and independently suppress apoptosis in our cell system through a different molecular mechanism. Although Hsp70 was reported to have an anti-apoptotic effect through veiled mechanism [23], the mechanism of such effect needs to be examined in further studies.

Finally, we highlighted that chaperones play a role in the development of polyQ disease. These evidences suggested that increasing expression level or enhancing the function of chaperones provide an avenue for the treatment of polyQ disease. Further studies of cellular and animal models are required to determine the precise mechanism of neurodegeneration of polyQ disease mediated by expanded polyglutamine tract as well as therapeutic approach.

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REFERENCES

1. Bruening, W.; Roy, J.; Giasson, B.; Figlewicz, D. A.; Mushynski, W. E.; Durham, H. D. Up-regulation of protein chaperones preserves viability of cells expressing toxic Cu/Zn-superoxide dismutase mutants associated with amyotrophic lateral sclerosis. J. Neurochem. 72:693–699; 1999.

- 2. Bukau, K. T.; Horwich, A. L. The Hsp70 and Hsp60 chaperone machines. Cell 92:351–366; 1998.
- 3. Burke, J. R.; Enghild, J. J.; Martin, M. E.; Jou, Y. S.; Myers, R. M.; Roses, A. D.; Vance, J. M.; Strittmatter, W. J. Huntingtin and DRPLA proteins selectively interact with the enzyme GAPDH. Nat. Med. 2:347–350; 1996.
- 4. Caplan, A. J.; Cyr, D. M.; Douglas, M. G. Eukaryotic homologues of Escherichia coli dnaJ: A diverse protein family that functions with hsp70 stress proteins. Mol. Biol. Cell 4:555–563; 1993.
- 5. Carmichael, J.; Chatellier, J.; Woolfson, A.; Milstein, C.; Fersht, A. R.; Rubinsztein, D. C. Bacterial and yeast chaperones reduce both aggregate formation and cell death in mammalian cell models of Huntington's disease. Proc. Natl. Acad. Sci. USA 97:9701–9705; 2000.
- 6. Chadli, A.; Bouhouche, I.; Sullivan, W.; Stensgard, B.; McMahon, N.; Catelli, M. G.; Toft, D. O. Dimerization and N-terminal domain proximity underlie the function of the molecular chaperone heat shock protein 90. Proc. Natl. Acad. Sci. USA 297:12524–12529; 2000.
- 7. Chai, Y.; Koppenhafer, S. L.; Bonini, N. M.; Paulson, H. L. Analysis of the role of heat shock protein (Hsp) molecular chaperones in polyglutamine disease. J. Neurosci. 19:10338–10347; 1999.
- 8. Chai, Y.; Koppenhafer, S. L.; Shoesmith, S. J.; Perez, M. K.; Paulson, H. L. Evidence for proteasome involvement in polyglutamine disease: Localization to nuclear inclusions in SCA3/MJD and suppression of polyglutamine aggregation *in vitro*. Hum. Mol. Genet. 8:673–682; 2000.
- 9. Chan, H. Y. E.; Warrick, J. M.; Gray-Board, G. L.; Paulson, H. L.; Bonini, N. M. Mechanisms of chaperone suppression of polyglutamine disease: Selectivity, synergy and modulation of protein solubility in *Drosophila*. Hum. Mol. Genet. 9:2811–2820; 2000.
- 10. Chellaiah, A.; Davis, A.; Mohanakumar, T. Cloning of a unique human homologue of the Escherichia coli DNAJ heat shock protein. Biochim. Biophys. Acta 1174:111–113; 1993.
- 11. Ciechanover, A. The ubiquitin-proteasome pathway: On protein death and cell life. EMBO J. 17:7151–7160; 1998.
- 12. Cummings, C. J.; Mancini, M. A.; Antalffy, B.; DeFranco, D. B.; Orr, H. T.; Zoghbi, H. Y. Chaperone suppression of aggregation and altered subcellular proteasome localization imply protein misfolding in SCA1. Nat. Genet. 19:148–154; 1998.
- 13. Davies, S. W.; Turmaine, M.; Cozens, B. A.; DiFiglia, M.; Sharp, A. H.; Ross, C. A.; Scherzinger, E.; Wanker, E. E.; Mangiarini, L.; Bates, G. P. Formation of neuronal intranuclear inclusions underlies the neurological dysfunction in mice transgenic for the HD mutation. Cell 90:537–548; 1997.
- 14. DiFiglia, M.; Snapp, E.; Chase, K. O.; Davies, S. W.; Bates, G. P.; Vonsattel, J. P.; Aronin, N. Aggregation of huntingtin in neuronal intranuclear inclusions and dystrophic neurites in brain. Science 227: 1990–1993; 1997.
- 15. Doyu, M.; Sobue, G.; Mukai, E.; Kachi, T.; Yasuda, T.; Mitsuma, T.; Takahashi, A. Severity of X-linked recessive bulbospinal neuronopathy correlates with size of the tandem CAG repeat in androgen receptor gene. Ann. Neurol. 23:707–710; 1992.
- 16. Fink, A. L.; Goto, Y. Molecular chaperones in the life cycle proteins. New York: Marcel Dekker, Inc.; 1998.
- 17. Goldberg, Y. P.; Nicholson, D. W.; Rasper, D. M.; Kalchman, M. A.; Koide, H. B.; Graham, R. K.; Bromm, M.; Kazemi-Esfarjani, P.; Thornberry, N. A.; Vaillancourt, J. P.; Hayden, M. R. Cleavage of huntingtin by apopain; a proapoptotic cysteine protease; is modulated by the polyglutamine tract. Nat. Genet. 13:442–449; 1996.
- 18. Gregori, L.; Fuchs, C.; Figueiredo-Pereira, M. E.; Nostrand, W. E. V.; Goldgaber, D. Amyloid beta-protein inhibits ubiquitin-dependent protein degradation in vitro. J. Biol. Chem. 270:19702–19708; 1995.
- 19. Hartl, F. U. Molecular chaperones in cellular protein folding. Nature 381:571–579; 1996.
- 20. Hendricks, J. P.; Hartl, F. U. Molecular chaperone functions of heatshock proteins. Annu. Rev. Biochem. 62:349–384; 1993.
- 21. Igarashi, S.; Koike, R.; Shimohata, T.; Yamada, M.; Hayashi, Y.; Takano, H.; Date, H.; Oyake, M.; Sato, T.; Sato, A.; Egawa, S.; Ikeuchi, T.; Tanaka, H.; Nakano, R.; Tanaka, K.; Hozumi, I.; Inuzuka, T.; Takahashi, H.; Tuji, S. Suppression of aggregate formation and

apoptosis by transglutaminase inhibitors in cells expressing truncated DRPLA protein with an expanded polyglutamine stretch. Nat. Genet. 18:111–117; 1998.

- 22. Ikeda, H.; Yamaguchi, M.; Sugai, S.; Aze, Y.; Narumiya, S.; Kakizuka, A. Expanded polyglutamine in the Machado-Joseph disease protein induces cell death in vitro and in vivo. Nat. Genet. 13:196– 202; 1996.
- 23. Jäättelä, M.; Wissing, D.; Kokholm, K.; Kallunki, T.; Egeblad, M. Hsp70 exerts its anti-apoptotic function downstream of caspase-3-like proteases. EMBO J. 17:6124–6134; 1998.
- 24. Jana, N. R.; Tanaka, M.; Wang, G. H.; Nukina, N. Polyglutamine length-dependent interaction of Hsp40 and Hsp70 family chaperones with truncated N-terminal huntingtin: Their role in suppression of aggregation and cellular toxicity. Hum. Mol. Genet. 9:2009–2018; 2000.
- 25. Kahlem, P.; Terre, C.; Green, H.; Djian, P. Peptides containing glutamine repeats as substrates for transglutaminase-catalyzed cross-linking: Relevance to diseases of the nervous system. Proc. Natl. Acad. Sci. USA 93:14580–14585; 1996.
- 26. Kalchman, M. A.; Koide, H. B.; McCutcheon, K.; Graham, R. K.; Nichol, K.; Nishiyama, K.; Kazemi-Esfarjani, P.; Lynn, F. C.; Wellington, C.; Metzler, M.; Goldberg, Y. P.; Kanazawa, I.; Gietz, R. D.; Hayden, M. R. HIP1; a human homologue of S. cerevisiae Sla2p; interacts with membrane-associated huntingtin in the brain. Nat. Genet. 16:44–53; 1997.
- 27. Kawaguchi, Y.; Okamoto, T.; Taniwaki, M.; Aizawa, M.; Inoue, M.; Katayama, S.; Kawakami, H.; Nakamura, S.; Nisimura, M.; Akiguchi, I.; Kimura, J.; Narumiya, S.; Kakizuka, A. CAG expansions in a novel gene for Machado-Joseph disease at chromosome 14q32.1. Nat. Genet. 8:221–227; 1994.
- 28. Klement, I. A.; Skinner, P. J.; Kaytor, M. D.; Yi, H.; Hersch, S. M.; Clark, H. B.; Zoghbi, H. Y.; Orr, H. T. Ataxin-1 nuclear localization and aggregation: Role in polyglutamine-induced disease in SCA1 transgenic mice. Cell 95:41–53; 1998.
- 29. Kobayashi, Y.; Kume, A.; Li, M.; Doyu, M.; Hata, M.; Ohtsuka, K.; Sobue, G. Chaperones Hsp70 and Hsp40 suppress aggregate formation and apoptosis in cultured neuronal cells expressing truncated androgen receptor protein with expanded polyglutamine tract. J. Biol. Chem. 275:8772–8778; 2000.
- 30. Kobayashi, Y.; Miwa, S.; Merry, D. E.; Kume, A.; Li, M.; Doyu, M.; Sobue, G. Caspase-3 cleaves the expanded androgen receptor protein of spinal and bulbar muscular atrophy in a polyglutamine repeat length-dependent manner. Biochem. Biophys. Res. Commun. 252: 145–150; 1998.
- 31. Koide, R.; Ikeuchi, T.; Onodera, O.; Tanaka, H.; Igarashi, S.; Endo, K.; Takahashi, H.; Kondo, R.; Ishikawa, A.; Hayashi, T.; Saito, M.; Tomoda, A.; Miike, T.; Naito, H.; Ikuta, F.; Tsuji, S. Unstable expansion of CAG repeat in hereditary dentatorubral-pallidoluysian atrophy (DRPLA). Nat. Genet. 6:9–13; 1994.
- 32. La Spada, A. R.; Roling, D. B.; Harding, A. E.; Warner, C. L.; Spiegel, R.; Hausmanowa-Petrusewicz, I.; Yee, W. C.; Fischbeck, K. H. Meiotic stability and genotype-phenotype correlation of the trinucleotide repeat in X-linked spinal and bulbar muscular atrophy. Nat. Genet. 2:301–304; 1992.
- 33. Li, M.; Miwa, S.; Kobayashi, Y.; Merry, D.; Tanaka, F.; Doyu, M.; Hashizume, Y.; Fischbeck, K. H.; Sobue, G. Nuclear inclusions of the androgen receptor protein in spinal and bulbar muscular atrophy. Ann. Neurol. 44:249–254; 1998.
- 34. Li, M.; Nakagomi, Y.; Kobayashi, Y.; Merry, D. E.; Tanaka, F.; Doyu, M.; Mitsuma, T.; Fischbeck, K. H.; Sobue, G. Nonneural nuclear inclusions of androgen receptor protein in spinal and bulbar muscular atrophy. Am. J. Pathol. 153:695–701; 1998.
- 35. Li, X. J.; Li, S. H.; Sharp, A. H.; Nucifora, F. C. Jr.; Schilling, G.; Lanahan, A.; Worley, P.; Snyder, S. H.; Ross, C. A. A huntingtinassociated protein enriched in brain with implications for pathology. Nature 378:398–402; 1995.
- 36. MacRae, T. H. Structure and function of small heat shock/alphacrystallin proteins: Established concepts and emerging ideas. Cell. Mol. Life Sci. 57:899–913; 2000.
- 37. Matilla, A.; Koshy, B. T.; Cummings, C. J.; Isobe, T.; Orr, H. T.; Zoghbi, H. Y. The cerebellar leucine-rich acidic nuclear protein interacts with ataxin-1. Nature 389:974–978; 1997.
- 38. Merry, D. E.; Kobayashi, Y.; Bailey, C. K.; Taye, A. A.; Fischbeck, K. H. Cleavage; aggregation and toxicity of the expanded androgen receptor in spinal and bulbar muscular atrophy. Hum. Mol. Genet. 7:693–701; 1998.
- 39. Michels, A. A.; Kanon, B.; Konings, A. W.; Ohtsuka, K.; Bensaude, O.; Kampinga, H. H. Hsp70 and Hsp40 chaperone activities in the cytoplasm and the nucleus of mammalian cells. J. Biol. Chem. 272: 33283–33289; 1997.
- 40. Minami, Y.; Hohfeld, J.; Ohtsuka, K.; Hartl, F. U. Regulation of the heat-shock protein 70 reaction cycle by the mammalian DnaJ homolog; Hsp40. J. Biol. Chem. 271:19617–19624; 1996.
- 41. Nagafuchi, S.; Yanagisawa, H.; Sato, K.; Shirayama, T.; Ohsaki, E.; Bundo, M.; Takeda, T.; Tadokoro, K.; Kondo, I.; Murayama, N.; Tanaka, Y.; Kikushima, H.; Umino, K.; Kurosawa, H.; Furukawa, T.; Nihei, K.; Inoue, T.; Sano, A.; Komura, O.; Takahashi, M.; Yoshizawa, T.; Kanazawa, I.; Yamada, M. Dentatorubral and pallidoluysian atrophy expansion of an unstable CAG trinucleotide on chromosome 12p. Nat. Genet. 6:14–18; 1994.
- 42. Oh, S.; Iwahori, A.; Kato, S. Human cDNA encoding DnaJ protein homologue. Biochim. Biophys. Acta 1174:114–116; 1993.
- 43. Paulson, H. L.; Perez, D. M. K.; Troffier, Y.; Trojanowski, J. Q.; Subramony, S. H.; Das, S. S.; Vig, P.; Mandel, J.-L.; Fischbeck, K. H.; Pittman, R. N. Intranuclear inclusions of expanded polyglutamine protein in spinocerebellar ataxia type 3. Neuron 19:333–344; 1997.
- 44. Perutz, M. F.; Johnson, T.; Suzuki, M.; Finch, J. T. Glutamine repeats as polar zippers: Their possible role in inherited neurodegenerative diseases. Proc. Natl. Acad. Sci. USA 91:5355–5358; 1994.
- 45. Saudou, F.; Finkbeiner, S.; Devys, D.; Greenberg, M. E. Huntingtin acts in the nucleus to induce apoptosis but death does not correlate with the formation of intranuclear inclusions. Cell 95:55–66; 1998.
- 46. Shimohata, T.; Nakajima, T.; Yamada, M.; Uchida, C.; Onodera, O.; Naruse, S.; Kimura, T.; Koide, R.; Nozaki, K.; Sano, Y.; Ishiguro, H.; Sakoe, K.; Ooshima, T.; Sato, A.; Ikeuchi, T.; Oyake, M.; Sato, T.; Aoyagi, Y.; Hozumi, I.; Nagatsu, T.; Takiyama, Y.; Nishizawa, M.; Goto, J.; Kanazawa, I.; Davidson, I.; Tanese, N.; Tsuji S. Expanded polyglutamine stretches interact with TAFII130; interfering with CREB-dependent transcription. Nat. Genet. 26:29–36; 2000.
- 47. Skinner, P. J.; Koshy, B. T.; Cummings, C. J.; Klement, I. A.; Helin, K.; Servadio, A.; Zoghbi, H. Y.; Orr, H. T. Ataxin-1 with an expanded glutamine tract alters nuclear matrix-associated structures. Nature 389: 971–974; 1997.
- 48. Stenoien, D. L.; Cummings, C. J.; Adams, H. P.; Mancini, M. G.; Patel, K.; DeMartino, G. N.; Marcelli, M.; Weigel, N. L.; Mancini, M. A. Polyglutamine-expanded androgen receptors form aggregates that sequester heat shock proteins; proteasome components and SRC-1; and are suppressed by the HDJ-2 chaperone. Hum. Mol. Genet. 8:731–741; 1999.
- 49. Stott, K.; Backburn, J. M.; Butler, P. J. G.; Perutz, M. Incorporation of glutamine repeats makes protein oligomerize: Implications for neurodegenerative diseases. Proc. Natl. Acad. Sci. USA 92:6509–6513; 1995.
- 50. The Huntington's Disease Collaborative Research Group. A novel gene containing a trinucleotide repeat that is expanded and unstable on Huntington's disease chromosomes. The Huntington's Disease Collaborative Research Group. Cell 72:971–983; 1993.
- 51. Waragai, M.; Lammers, C.; Takeuchi, S.; Imafuku, I.; Udagawa, Y.; Kanazawa, I.; Kawabata, M.; Mouradian, M. M.; Okazawa, H. PQBP-1; a novel polyglutamine tract-binding protein; inhibits transcription activation by Brn-2 and affects cell survival. Hum. Mol. Genet. 8:977–987; 1999.
- 52. Warrick, J. M.; Paulson, H. L.; Gray-Board, G. L.; Bui, Q. T.; Fischbeck, K. H.; Pittman, R. N.; Bonini, N. M. Expanded polyglutamine protein forms nuclear inclusions and causes neural degeneration in *Drosophila*. Cell 93:939–949; 1998.
- 53. Yamagishi, N.; Nishihori, H.; Ishihara, K.; Ohtsuka, K.; Hatayama, T. Modulation of the chaperone activities of Hsc70/Hsp40 by Hsp105alpha and Hsp105beta. Biochem. Biophys. Res. Commun. 272:850–855; 2000.
- 54. Yvert, G.; Lindenberg, K. S.; Picaud, S.; Landwehrmeyer, G. B.; Sahel, J. A.; Mandel, J. L. Expanded polyglutamines induce neurodegeneration and trans-neuronal alterations in cerebellum and retina of SCA7 transgenic mice. Hum. Mol. Genet. 9:2491–2506; 2000.
- 55. Zoghbi, H. Y.; Orr, H. T. Glutamine repeats and neurodegeneration. Annu. Rev. Neurosci. 23:217–247; 2000.